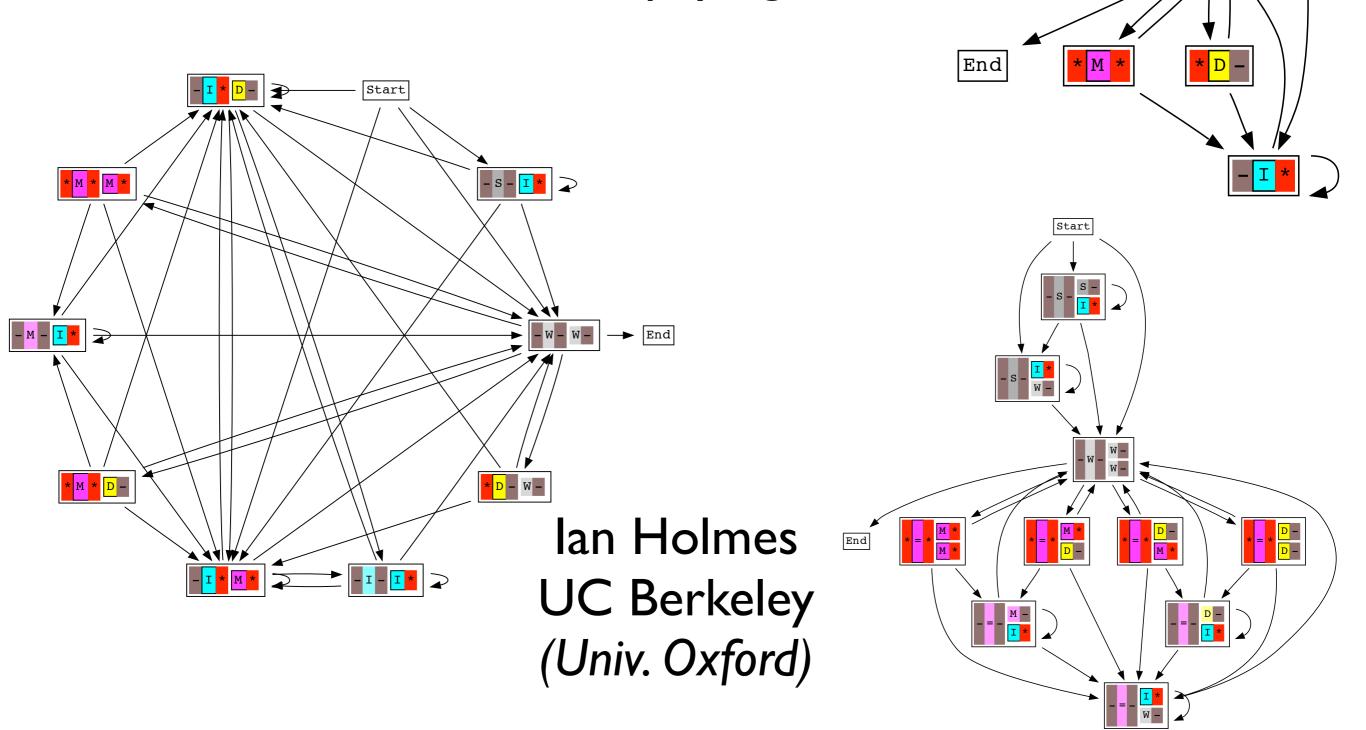
Workshop Announcement

- "Inference in Sequence Evolution"
- Mathematical Biosciences Institute, Ohio State University
- First quarter of 2010 (exact date TBA)
- Organizers: I.Holmes & G.Lunter

Transducers

A probabilistic framework for modeling insertions & deletions on phylogenetic trees

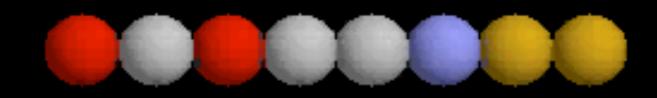


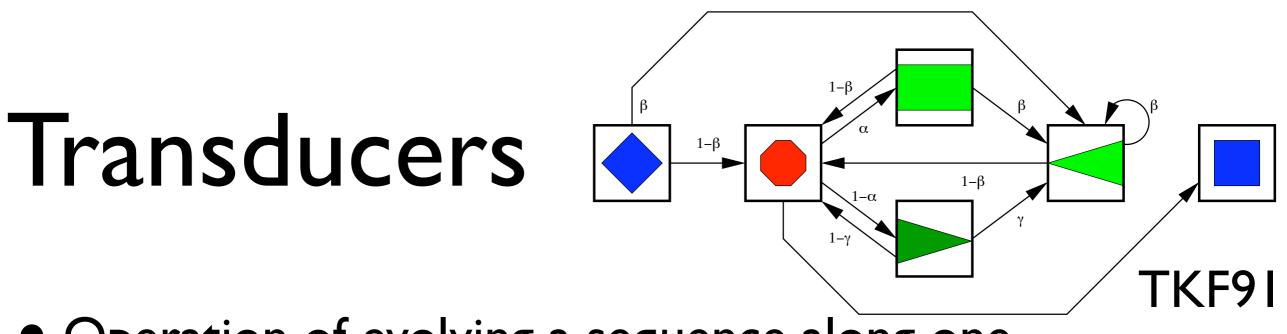
Start

W

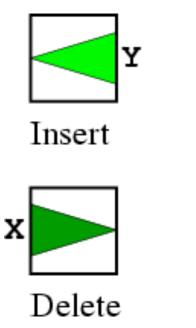
Phylo-Alignment

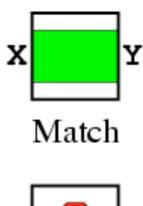
Phylo-Alignment





- Operation of evolving a sequence along one branch of a phylogenetic tree
- Represent as a finite state machine
- Input "tape" X, output "tape" Y



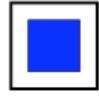




Wait



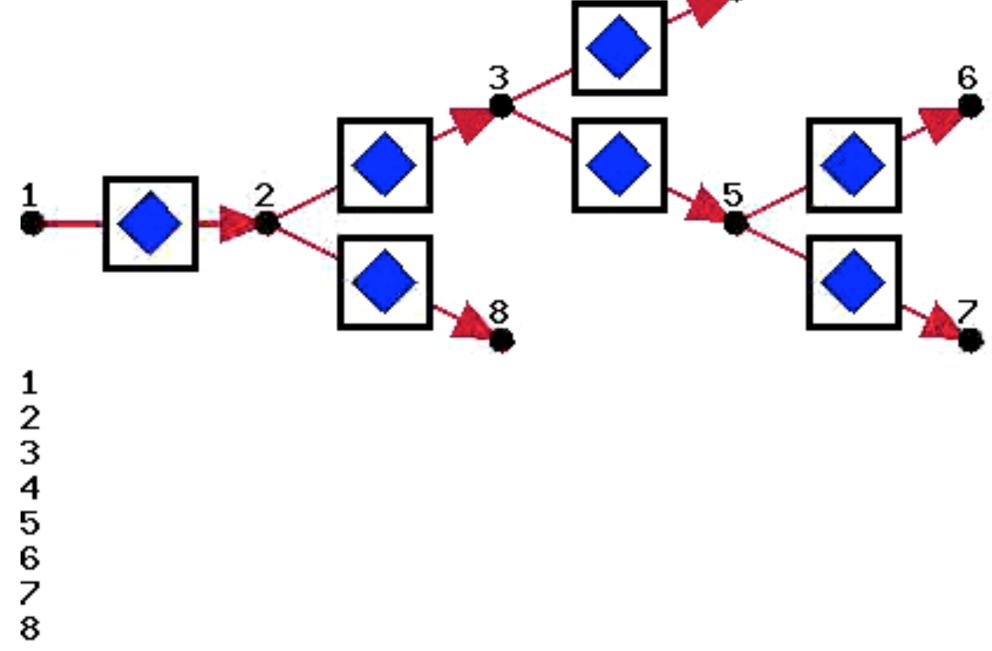
Start



End

Pair HMM: P(X,Y) Transducer: P(Y|X)

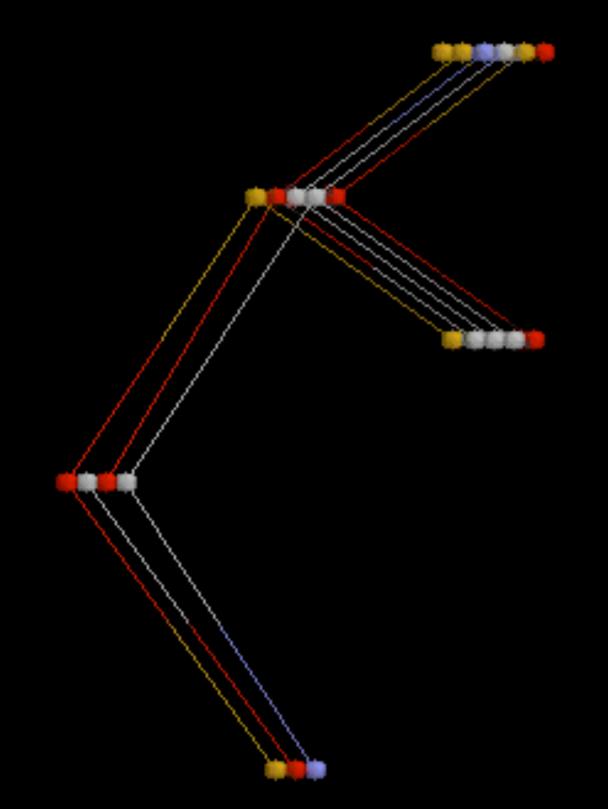
String transducers on a tree 3



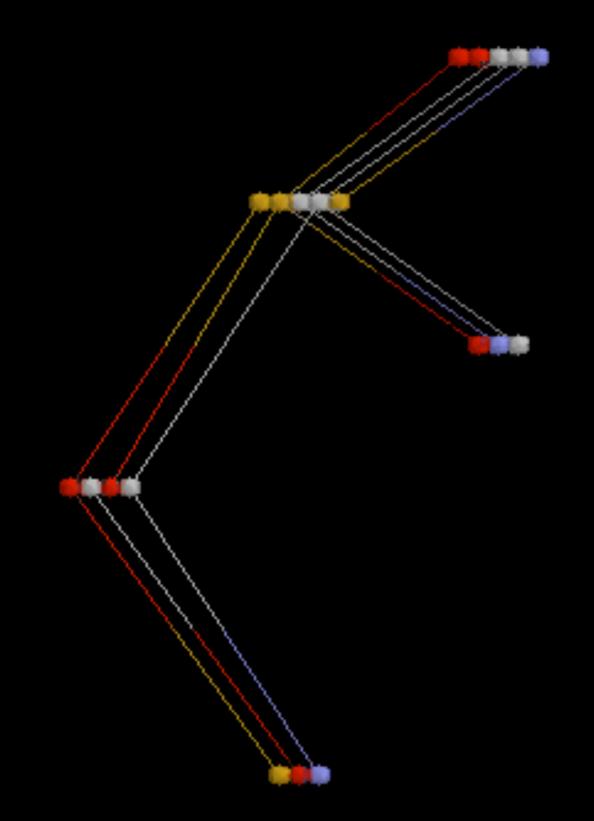
Mohri, Computational Linguistics, 1997 Hein, PSB, 2001; Holmes & Bruno, Bioinf., 2001 Holmes, Bioinformatics, 2003 also Bishop & Thompson; Thorne, Kishino & Felsenstein; Steel; Lunter, Miklos; Kim & Sinha; Satija, Pachter; Paten; Haussler ...

Curse of dimensionality

- Number of states in composite transducer increases exponentially with number of taxa
- So does the number of cells in the DP matrix
- Solution: Markov Chain Monte Carlo
 - Hold some parts of the state path fixed, while resampling other parts (Gibbs sampling)



Sampling one branch at a time



Sampling one node at a time

tkfalign vs Clustal (BAliBase)

BAliBASE subcategory	Prog.	Ref.	Iter.	CLUSTALW
Equidistant, similar lengths; high ID ($>35\%$)	0.775	0.784	0.774	0.884
Equidistant, similar lengths; medium ID $(20\% - 40\%)$	0.673	0.689	0.693	0.790
Equidistant, similar lengths; low ID ($< 25\%$)	0.654	0.658	0.669	0.787
Close family (> 25%) plus "orphan" outliers (< 20%)	0.814	0.827	0.839	0.928
Divergent subfamilies ($< 20\%$ between subfamilies)	0.481	0.525	0.528	0.693
Long gaps at the ends: N/C terminal extensions	0.348	0.359	0.372	0.672
Long gaps in the middle: Insertions	0.573	0.603	0.622	0.789

- Prog. = Progressive alignment
- Ref. = Refinement
- Iter. = MCMC + refinement

Holmes & Bruno, 2001

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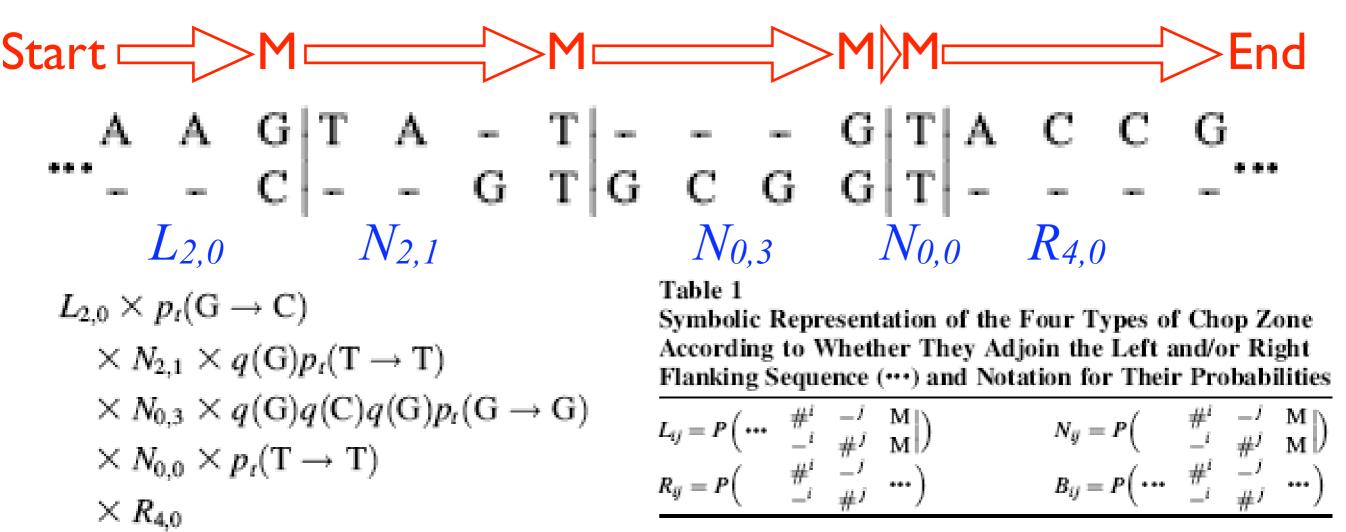
The "Long Indel" Model

Deletion events are "attached" to the leftmost residue that they remove. If residue N survives, then residue N+1 is unaffected by deletion events from the left

N N+I

Miklòs, Lunter & Holmes, 2004

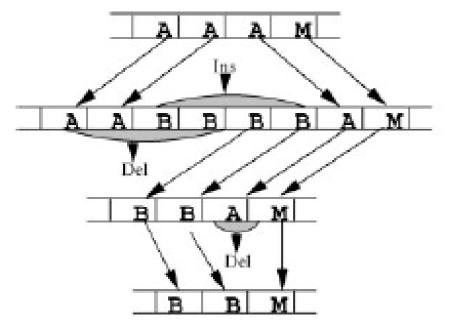
Independent "chop zones"



NOTE.—These probabilities are conditional on observing the i (or i + 1) ancestral nucleotides. The # signs represent unaligned residues; M pairs represent aligned residues, and vertical bars represent chop zone boundaries.

Miklòs, Lunter & Holmes, 2004

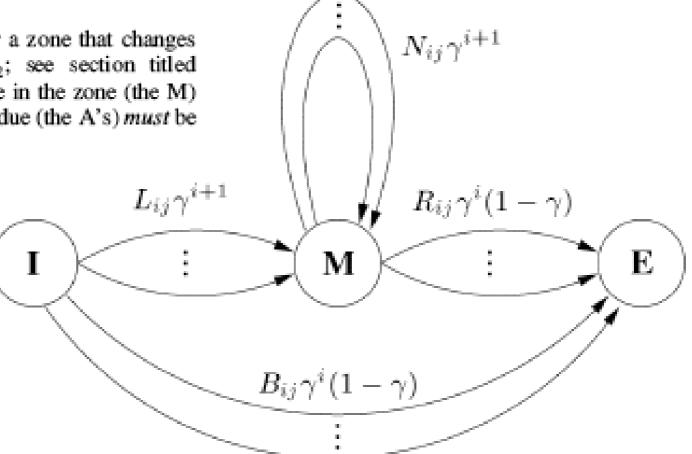
Generalized Pair HMM



4-residue insertion 4-residue deletion Gap penalties calculated by direct enumeration of trajectories (shown left: *N*_{3,2})

1-residue deletion

FIG. 1.—An example three-event trajectory for a zone that changes length from four resides to three (outcome $B_{3,2}^i$; see section titled *Algorithm*). By definition, the final ancestral residue in the zone (the M) cannot be deleted, whereas every other ancestral residue (the A's) *must* be deleted.



Shown right: the generalized Pair HMM for the "long indel" model

FIG. 3.—A hidden Markov model formulation of the long indel model. The emission probabilities (associated to transitions) are not included. The parameter $\gamma = \lambda_1/\mu_1$ is the parameter governing the geometric equilibrium length distribution.

Long Indel vs Gotoh (Homstrad)

Table 3Performance of Alignment Methods, as Measured byAlignment Accuracy or "Overlap," the Percentage ofAlignment Columns Identical to Those of the HOMSTRADStructural Alignments

Alignment Method	Training Set Optimization ^a	Test Set Overlap (%)
TKF91	ML	73.8
TKF92	ML	75.9
Gotoh (BLOSUM62)	NCBI defaults	80.9
Long indel	ML	81.1
Long indel, mixed geometric	Accuracy	82.1
Gotoh (BLOSUM62)	Accuracy	82.2

^a Parameters were optimized over a training set to maximize either likelihood or overlap. In addition, for the Gotoh algorithm we used NCBI (National Center for Biotechnology Information) defaults for gap opening and gap extension parameters.

Miklòs, Lunter & Holmes, 2004

"Every good work of software starts by scratching a developer's personal itch" - Eric Raymond

<u>2001</u> (Holmes & Bruno) MCMC based on TKF91. **Poor performance due to affine gaps, rate variation**

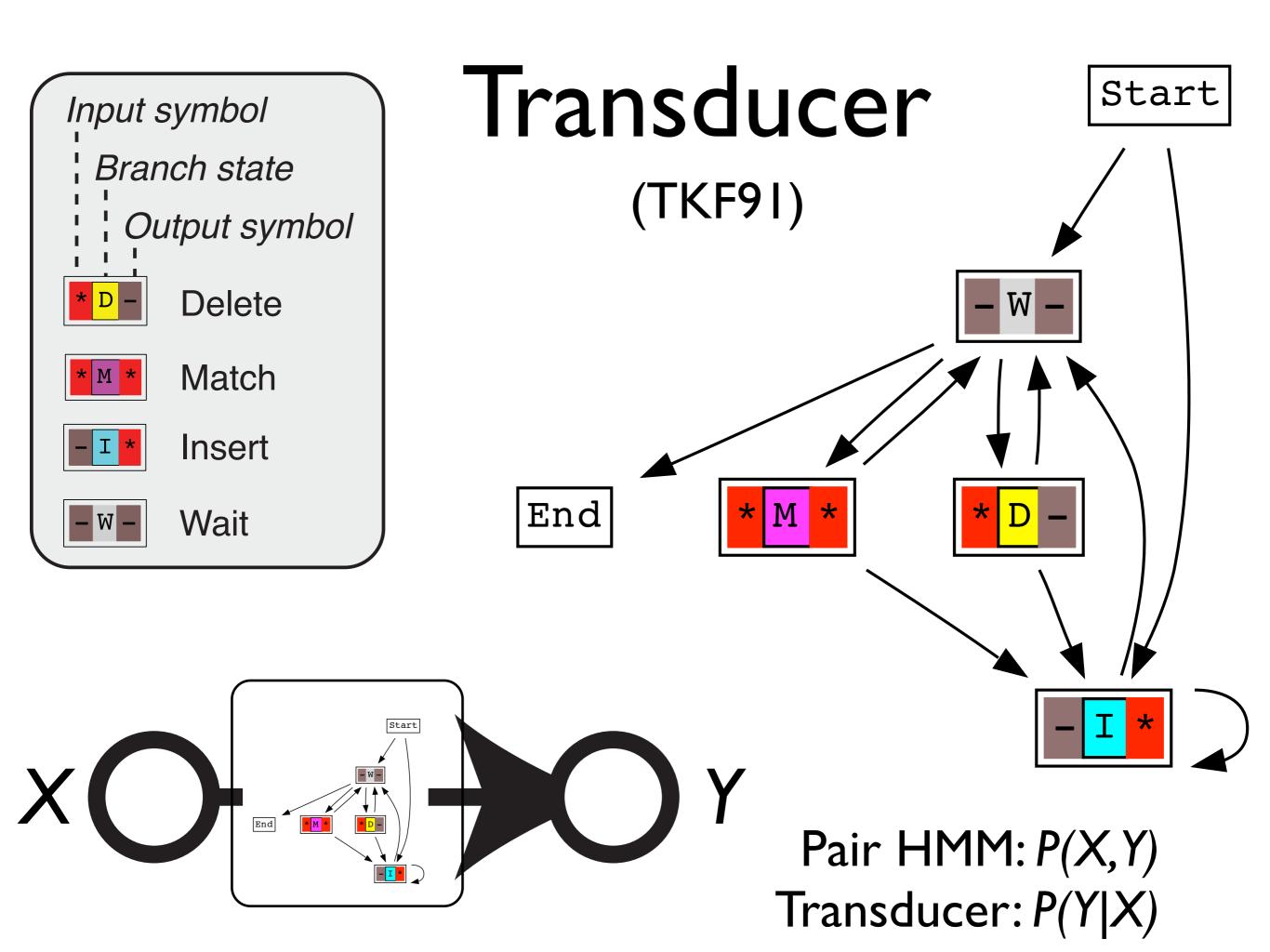
<u>2002</u> (Holmes & Rubin) EM algorithm for estimating substitution rates & in particular rate variation

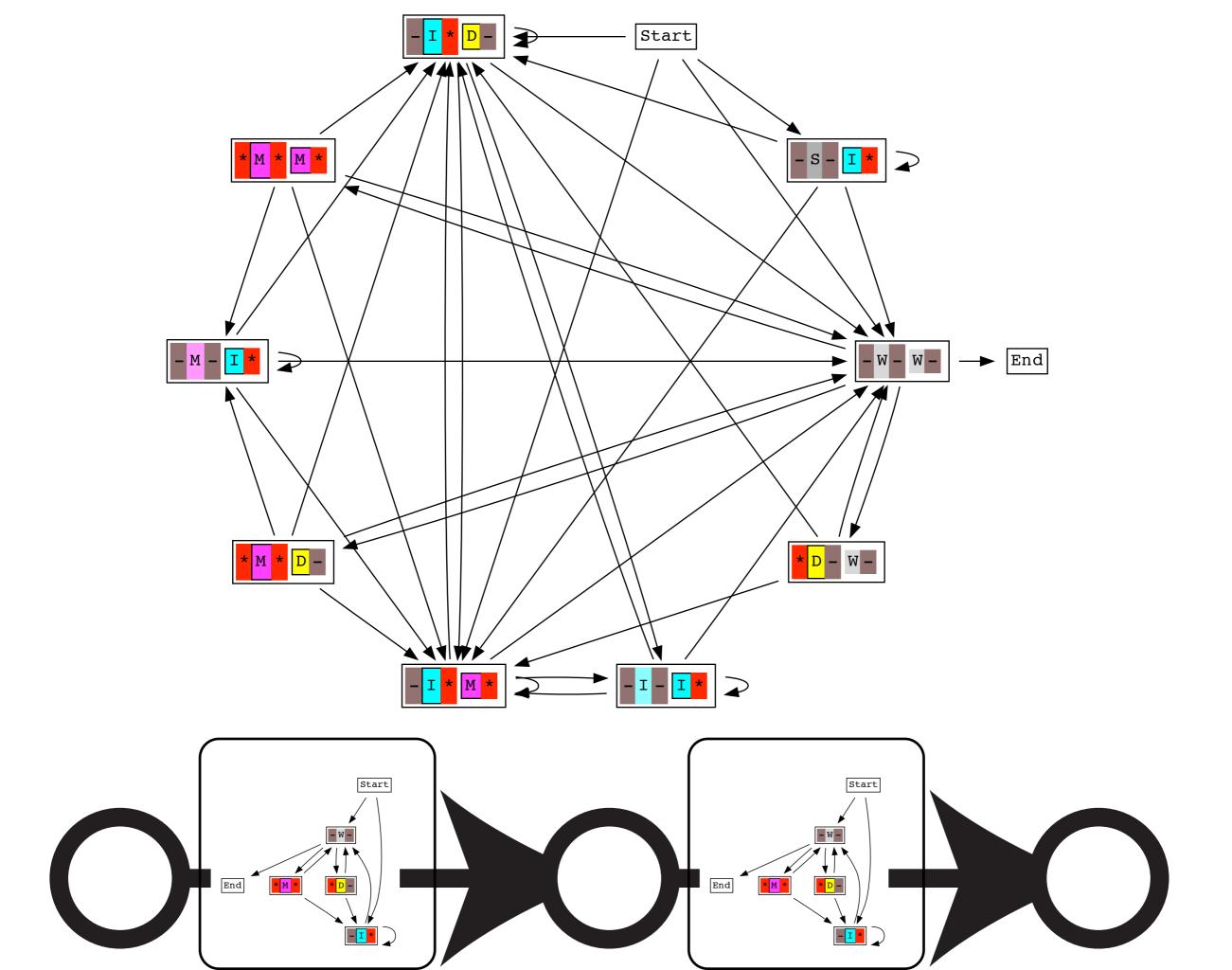
<u>2003</u> (Holmes) General algorithm for transducer composition on phylogenetic trees

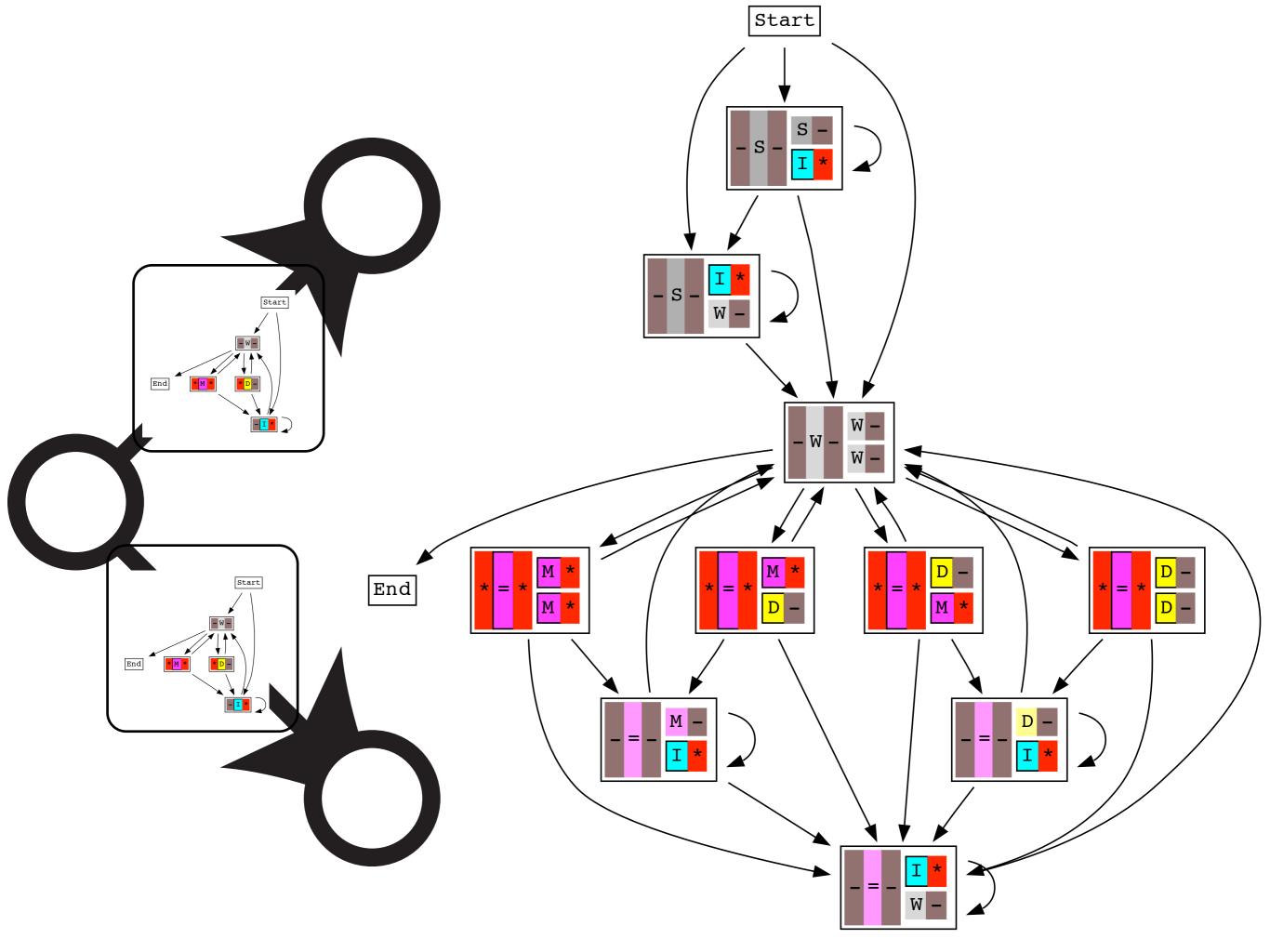
<u>2004</u> (Miklòs, Lunter & Holmes) "Long Indel" model: affine-gap Pair HMMs from evolutionary models

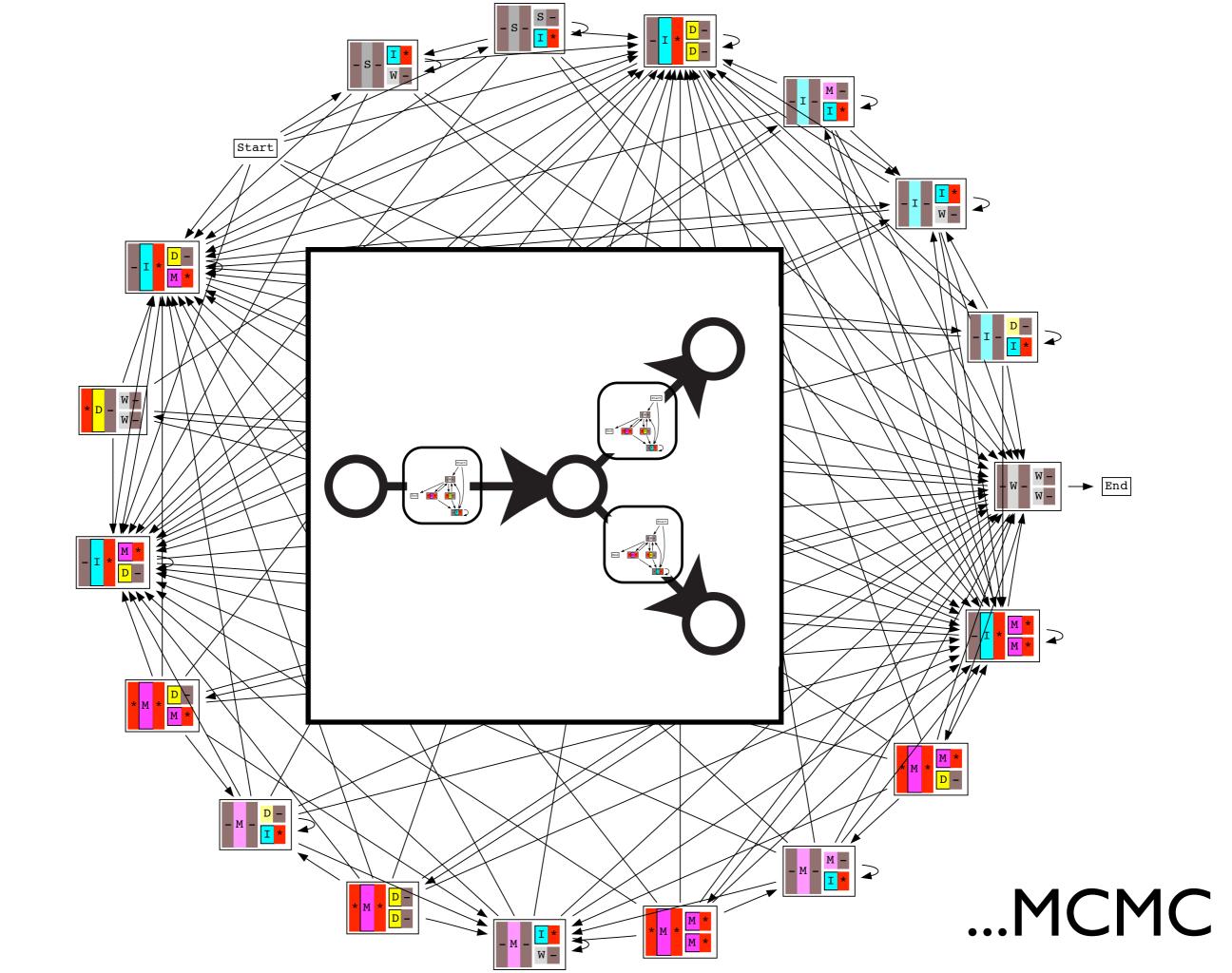
<u>2005-2006</u> (Holmes; Klosterman et al) (Started extending transducer theory to RNA sequence analysis & SCFGs) Enormous difficulties debugging transducer composition & sampling algorithms!

2007 (Holmes) Phylocomposer

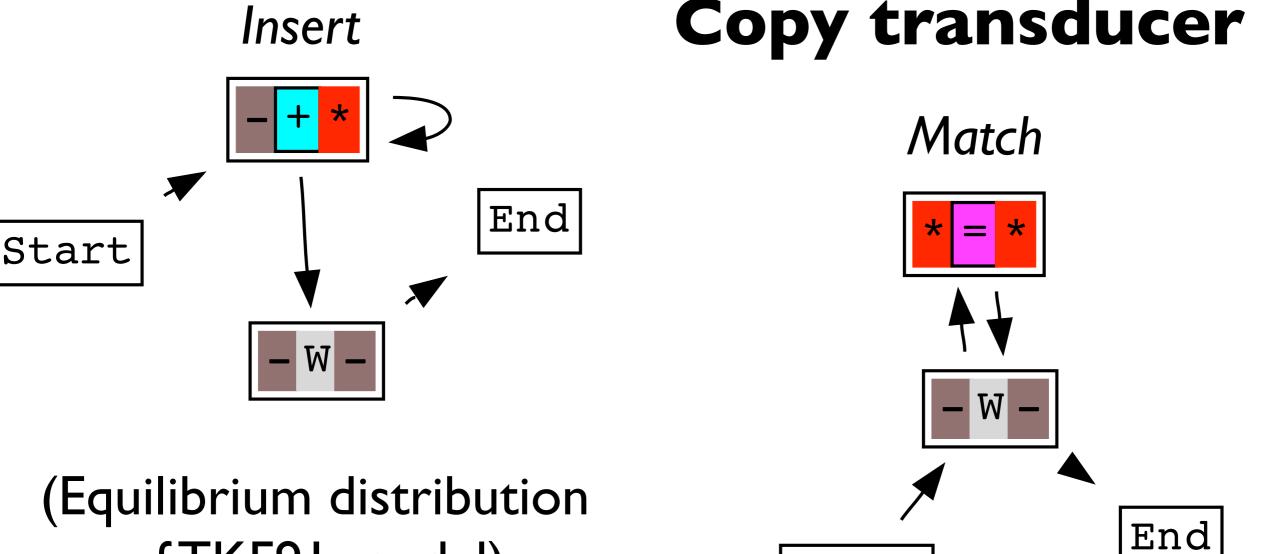








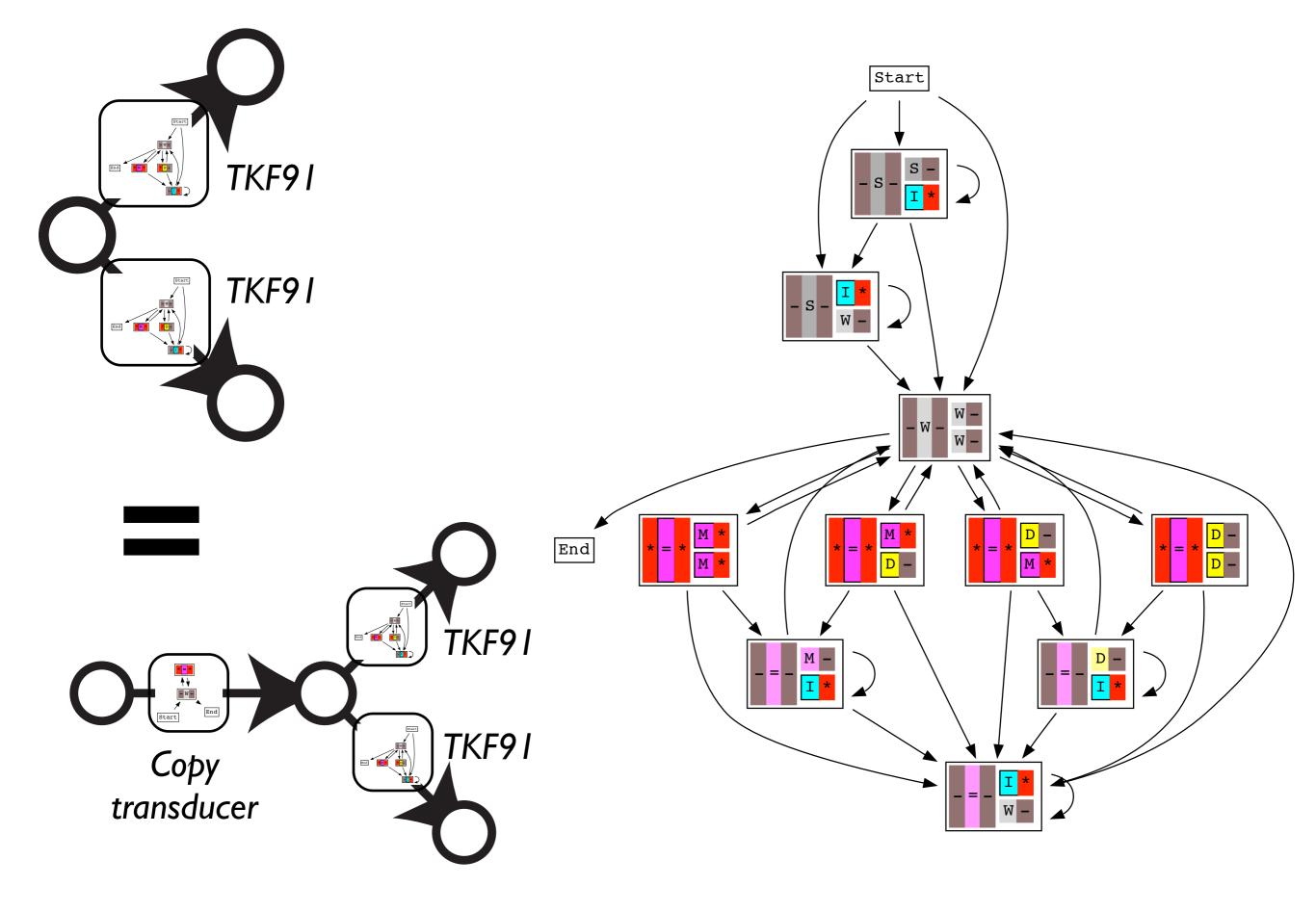
Singleton transducer



of TKF91 model)

(Zero-length branch in TKF91 model)

Start



HMMoC adapter

Calmes'

Gerton Lunter's HMMoC Hidden Markov Model Compiler

Speedup factor 10²-10³

Handel MCMC moves

Try a change in tree topology (*aunt attack*); sample alignment ABCD for both configurations tkfalign handalign

A B B Sample t(AB) (branch scaling)

Sample alignment AB (branch realignment)

Sample alignment ABCD and sequence at B (node realignment) A B D Sample t(AB) & t(BD), keeping the sum constant (branch sliding)

MCMC Statistical Alignment

